

Department of Soil, Water & Environmental Science

Evaluation of 3 Clearbrook Portable Water Purification Units for the Removal of *Cryptosporidium parvum* oocysts & *Escherichia coli*

SUMMARY

Three Clearbrook Portable (Mobile, AL) Water Purification Units (drinking bottle type) were evaluated for their ability to remove *Cryptosporidium parvum* (*C. parvum*) oocysts, and *Escherichia coli* (*E. coli*). The average oocysts removal by both units tested exceeded >99.99%, whereas the removal of *E. coli* was >99.99999%.

METHODOLOGY

Test Waters and Test Conditions

Typical chemical/physical parameters of the test water used in the study are shown in Table 1. This water was within the range recommended by the U.S. Environmental Protection Agency's Task Force Report on *Guide Standard and Protocol for Testing Microbiological Water Purifiers* (1990).

Waters for conditioning and testing were passed through the unit by negative pressure, with an Expert peristaltic pump (SciLog, Verona, WI), at an approximate flow rate of 200 ml/min. The test was conducted after conditioning the unit with approximately 4 liters of dechlorinated tap water. The challenge water (influent) contained approximately 1.4×10^7 *C. parvum* oocysts/L, and 1×10^8 colony forming units (CFU) of *E. coli*/L. Two influent samples (50 mL each) were collected at the beginning of the experiment, and two effluent samples (500 mL each) after passing approximately one and 1.8 liters of challenge water, respectively.

C. parvum Assay

C. parvum oocysts were obtained from feces of infected calves (Pleasant Hill Farm, Troy, ID). Oocysts were pelleted by centrifugation, and the supernatant was aspirated to approximately 1 mL above the pellet. After resuspension of the pellet in phosphate buffer saline, the oocysts were counted using a hemacytometer (Baxter Healthcare Corp. McGraw Park, IL). A total of 12 chamber aliquots were counted for each sample according to the procedure outlined in the Guidance Manual (USEPA, 1990). Influent and effluent samples were assayed in triplicate.

Bacterial Analysis

E. coli (ATCC-25922) was grown overnight in Trypticase Soy broth (Difco, Detroit, MI) at 37 degrees C to obtain the organisms in the stationary growth phase. The bacterial cells were pelleted by centrifugation and resuspended in phosphate buffered saline. This procedure was repeated three times to remove organic matter present in the broth. Bacterial assays were conducted by the membrane filtration method on m-Endo Agar LES (Becton Dickinson, Cookesville, MD). Appropriate dilutions of influent samples were made in sterile 0.025 M phosphate buffered saline (PBS) at pH 7.0. One, 10 and 100 mL volumes of effluent samples were assayed. All samples were assayed in triplicate.

RESULTS

The results of *C. parvum* oocysts and *E. coli* removal are shown in Table 2. These results show that the unit achieved an average removal of more than 4 log (>99.993%) of *C. parvum* oocysts, and more than 7 log (>99.99999%) of *E. coli*.

REFERENCES

USEPA 1990. Guide Standard and Protocol for Testing Microbiological Water Purifiers. In: Guidance Manual for Compliance with the filtration and Disinfection Requirements for Public Water Systems Using Surface Water Sources.

Table 1. Typical characteristics of non-microbiological parameters of test water

<u>Parameter</u>	<u>Test water</u>
Chlorine residual (ppm)	<0.1
pH	7.7
Turbidity (NTU)	0.22
Temperature (C.)	24.5
Total dissolved solids (TDS) (mg/L)	221
Total organic carbon (TOC) (mg/L)	ND

Table 2. Removal of *C. parvum* oocysts and *E. coli* by three Clearbrook portable water purification units

<u>Organism</u>	<u>Influent</u>	<u>Effluent</u>	<u>% reduction</u>	<u>Log reduction</u>
<i>C. parvum</i>	1.4×10^7	$< 9.2 \times 10^{-2}$	> 99.993	> 4.2
<i>E. coli</i>	1.0×10^8	< 10	> 99.99999	> 7.0